



CHARACTERIZATION AND PYROLYSIS OF POST-CONSUMER LEATHER SHOE WASTE FOR THE RECOVERY OF VALUABLE **CHEMICALS**

Melissa Lisa Van Rensburg^{1,*}, S'phumelele Lucky Nkomo¹ and Ntandoyenkosi Malusi Mkhize²

¹ University of KwaZulu-Natal, Discipline of Geography, 238 Mazisi Kunene Road, Glenwood, 4041 Durban, KwaZulu Natal, South Africa ² University of KwaZulu-Natal, Discipline of Chemical Engineering, 238 Mazisi Kunene Road, Glenwood, 4041 Durban, KwaZulu-Natal, South Africa

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ABSTRACT

Majority of post-consumer leather footwear currently ends up in landfill sites with adverse environmental impacts. Current waste recovery options have proven largely unsuccessful in minimizing this waste stream. This study investigates whether leather from post-consumer footwear can be pyrolyzed using gram-scale (fixed-bed) and microgram-scale (TGA) pyrolysis reactors. The investigation was conducted using final pyrolysis process temperatures between 450 and 650 °C and solid residence times of 5 to 15 minutes. The purpose of the experiments was to assess the waste recovery potential of leather pyrolysis products for valuable chemicals. The pyrolysis product fractions (solid, liquid, and gas) distribution were investigated, optimal pyrolysis conditions presented, and the product fractions characterized for their elemental and chemical composition using ultimate and GC-MS analysis. The distribution of the product fractions proved leather footwear pyrolysis was viable under the given conditions. The completion of leather footwear pyrolysis was evident at 650°C since the solid yield reached a constant value of approximately 25 wt.%. The liquid fraction was maximized within the temperature range of 550-650°C (Max= 54 wt.%), suggesting optimal pyrolysis conditions within this range. The higher heating values (HHVs) of the pyrolysis leather oil (33.6 MJ/kg) and char (25.6 MJ/kg) suggested their potential application for energy or fuel. The liquid fraction comprised predominantly of nitrogen derivatives and potential applications areas include use in the production of fertilizers, chemical feedstocks, or the pharmaceutical industry. This study proved that leather from post-consumer footwear can be pyrolyzed and provided valuable insight into its characterization and potential applications areas.

1. INTRODUCTION

Post-consumer shoe waste is a fundamental problem facing the footwear industry and waste management sectors. The footwear industry is rapidly growing, and it was estimated that in 2017 the production of shoes reached 23.5 billion pairs (Portuguese Shoes, 2018). The rapid production and consumption of footwear correspond to a substantial amount of post-consumer waste, the majority of which are currently disposed of in landfill sites (Kolomaznik et al., 2008, and Chowdhury, 2018). According to Mia et al. (2017), the most significant environmental challenge facing the footwear industry is the large amount of post-consumer waste generated in the End-of-Life (EoL) phase.

Leather waste constitutes a significant portion of waste emanating from the footwear industry because it is

tuting approximately 25% leather by weight (Staikos et al., 2006). Leather waste from post-consumer shoes is detrimental in its EoL phase because of the adverse environmental impacts associated with its disposal. Studies detailing the negative environmental impacts of leather shoe disposal highlight issues related to freshwater contamination, eutrophication, degradation of ecosystem quality, harmful emissions, and Global Warming Potential (GWP) (Albers et al., 2008, Joseph and Nithya, 2009, and Gottfridson and Zhang, 2015). Additionally, the footwear industry accounts for 1.4% of global climate impacts (Chrobot et al., 2018:18), and it is estimated that a single pair of leather shoes takes approximately 25-40 years to decompose in landfill sites (Fyvie, 2018). This study examines the poten-

a prominent material used, with the average shoe consti-



tial of pyrolysis as a waste recovery avenue to reduce the amount of post-consumer leather shoe waste disposed of in landfill sites.

According to Czajczyńska et al. (2017), pyrolysis is the thermochemical devolatilization of organic or inorganic materials at elevated temperatures in an inert atmosphere. The heating of materials through pyrolysis leads to the formation of solid, liquid, and gas fractions (Arabiourrutia et al., 2020). Pyrolysis is advantageous over other waste treatment processes such as incineration because the oxygen-free atmosphere in which pyrolysis takes place causes fewer CO₂ emissions, thus lowering the greenhouse effect (Jo et al., 2017). Moreover, incineration and open combustion are inefficient waste recovery methods as the technology merely recovers energy while reducing the remainder of the materials to ash (Aylón et al., 2010). Murugan et al. (2008) state that pyrolysis technology can process the materials into three main fractions (solid, liquid, and gas) to recover valuable chemicals that would otherwise be lost during incineration or open combustion. These chemicals can be further processed into valuable products and hence improve waste recovery efficiency from materials. Food waste (Joe et al., 2017), plastic (Sorum et al., 2001, Olazar et al., 2009, and Sharuddin et al., 2016), and waste tyres (Islam et al., 2013, Perondi et al., 2016, and Lopez et al., 2017) are among some of the most widely studied materials using pyrolysis experiments. There are, however, limited studies that examine the potential of pyrolysis for recovering leather shoe waste.

Previous studies on leather pyrolysis focus mainly on waste from tanneries. A study on the production of useful materials from different types of leather tannery wastes was conducted by Yilmaz et al. (2007). The study investigated the experimental characteristics of leather waste pyrolysis at 450 and 600°C. The pyrolysis experiments indicated that leather waste heated using a fixed bed reactor at 450 and 600°C yielded liquid, gas, and char products. Increasing the final temperature from 450

TABLE 1: Summary of previous leather pyrolysis studies.

to 600°C caused the liquid yield to decrease from 32 to 28% and the gas yield to increase from 17 to 23%. The authors emphasized the desirability of activated carbon, a refined version of the pyrolysis solid fraction (char), from pyrolyzed leather waste. Similarly, a study by Sethuraman et al. (2013) subjected hazardous chrome tanned leather waste (CTLW) from tanneries to pyrolysis to obtain valuable products. The main useful products generated through this process included carbonaceous residual ash containing chromium, condensate oil, and fuel gas (Sethuramen et al., 2013).

Marcilla et al. (2012) investigated the effect of flash (450-550°C) and slow (up to 750°C) pyrolysis on bovine leather from tanneries. Flash pyrolysis generally involves a faster heating rate and a shorter solid residence time compared to slow pyrolysis (Al Arni, 2018). Authors indicated that the maximum liquid yield (42-45%) was achieved through flash pyrolysis between temperatures of 500-550°C, while slow pyrolysis presented significantly lower levels of liquid yield (29%). This finding is supported by literature indicating that flash pyrolysis generally optimizes the production of liquid yields while slow pyrolysis favors char as the main byproduct (Ronsse et al., 2012, and Al Arni, 2018). Kluska et al. (2019) focused on presenting the influence of temperature on the physicochemical properties of products obtained through the pyrolysis of leather tannery waste at lower temperature ranges. The leather waste was pyrolyzed at temperatures ranging from 300 to 500°C. The authors concluded that a maximum liquid yield of 42% was obtained at 400°C, followed by 40% at 500°C. A summary of the main leather waste pyrolysis studies and their corresponding yields is presented in Table 1.

Previous studies contribute to knowledge generation about the recovery of leather tannery waste, but they do not address the issue of post-consumer leather waste specifically for footwear products. It can thus be inferred that a gap in the literature exists to investigate whether leather shoe waste can be pyrolyzed similarly to tannery waste

Reference	Pyrolysis type	Temperature (°C)	Solid yield (wt.%)	Liquid yield (wt.%)	Gas yield (wt.%)	Sample mass (g)	Reactor	Solid residence time (min)
Sethuraman et al. (2013)	Slow	800	34.7	33.3	32.0	2000	Fixed bed	n.a
Yilmaz et al. (2007)	Slow	450	50.2	32.0	17.8	50-60	Fixed bed	120
(2007)		600	47.4	28.9	23.6	00 00		
Marcilla et al. (2012)	Fast	450	38.5	41.0	20.5	2	Fixed bed	n.a
()		500	30.4	44.5	25.1			
		550	31.0	42.3	26.6			
	Slow	700	28.6	29.6	41.8	4		
Kluska et al. (2019)	Slow	300	33.0	21.0	46.0		Fixed bed	30
(2013)		350	33.0	35.0	32.0			
		400	33.0	42.0	25.0	100		
		450	33.0	35.0	32.0			
		500	29.0	40.0	31.0			

to recover materials. This research is premised primarily as a concept proving and desirability study. The concept proving aspect is justified through the study's focus on investigating whether chrome tanned leather from post-consumer footwear has the potential to be pyrolyzed through experimental runs. The optimization of pyrolysis yields was also investigated through experimental runs, and this constituted the desirability aspect of the study. The novelty of the study reveals itself in the fact that leather from post-consumer footwear has yet to be pyrolyzed and hence motivates the need for this study as footwear presents challenges in the EoL phase.

The present study therefore examined waste recovery by thermochemical treatment by conducting pyrolysis experiments using samples of solid leather waste from post-consumer shoes. The main aim of this study was to assess the waste recovery potential of leather pyrolysis products for valuable chemicals. The aim was achieved through the following investigations (i) leather footwear pyrolysis experiments using fixed bed and TGA pyrolysis reactors; (ii) an analysis of the pyrolysis product distribution and optimal operating conditions; (iii) characterization of the elemental and chemical composition of the pyrolysis product yields using ultimate and GC-MS analysis, including an analysis of the higher heating value (HHV).

2. MATERIALS AND METHODS

2.1 Leather feedstock

Samples of leather shoes were obtained from various local shoe producers for pyrolysis. The samples consisted of reject shoes which are considered waste as they can no longer be used or sold. The shoe samples consisted of chrome tanned leather which was separated from the shoe soles to obtain leather in isolation from the other materials. The leather was cut into smaller pieces with particle sizes of approximately 0.5 cm. The shredded leather from various shoes was mixed before use in the experimental pyrol-

ysis runs to ensure feedstock homogeneity. The weight of each sample was 40g.

Preliminary experimental preparations included characterizing the sample material. To characterize and understand the constituents of the sample material, a proximate and ultimate analysis was conducted. The proximate analysis was performed to determine the major constituents of the feed material used in pyrolysis, while the ultimate analysis provided an elemental composition of the material (Basu, 2018). The results of the proximate and ultimate analyses of leather shoe waste are depicted in Table 2. The results are shown in comparison to previous studies which have been conducted on leather waste. These characteristics are essential to understand as they provide a broad understanding of the combustion characteristics of solid leather shoe waste.

2.2 Experimental setup

2.2.1 Fixed bed reactor

The pyrolysis unit chosen for this study was a fixed bed reactor. The fixed bed pyrolysis system comprised of a fixed bed of feedstock which was pyrolyzed as a batch reaction (Guda et al., 2015). The main features of the fixed bed reactor used in this study included a sample capacity of approximately ±45g (depending on the density and type of material) and a reactor temperature range between 0-1200°C. The dimensions of the fixed bed reactor can be found in Figure 1. The fixed bed reactor provided technology that was reliable and simple for the research purposes in this study (Aziz et al., 2018). A fixed bed reactor feedstock system was thus suitable for this research study as it was conducted on a small-scale for research purposes and allowed for a proof of the concept.

2.2.2 Pyrolysis system

The pyrolysis system used in this study is illustrated in Figure 1. This system involved the input of the leather waste feedstock into the sample holder placed inside of

TABLE 2: Ultimate and proximate analysis of leather waste in previous studies.

Reference	Sethuraman et al. (2013)	Yilmaz et al. (2007)	Marcilla et al. (2012)	Kluska et al. (2019)	Rodrigues et al. (2010)	Godinho et al. (2011)	Present study
Proximate analysis (wt. %)	•						
Moisture content	5.50	7.10	11.20	35.30	12.40	14.10	12.56
Volatile matter	60.30	67.00	76.70	67.90	67.70	66.40	61.23
Fixed carbon	26.62	n.d	6.12	21.20	14.80	14.52	18.77
Ash	7.58	9.60	6.03	10.90	5.10	4.95	7.44
Ultimate analysis (wt. %) ª			·		·		
Carbon	55.31	44.30	41.50	41.71	50.61	42.36	38.93
Hydrogen	7.86	3.10	6.85	7.12	8.79	7.32	5.62
Oxygen ^b	8.54	36.60	37.90	28.46	25.46	19.00	32.36
Nitrogen	12.56	14.20	12.90	11.01	12.80	10.67	8.88
-	1 69	1 80	0.83	3.43	1.89	1.57	1.65



FIGURE 1: Pyrolysis system used in this study.

the fixed bed reactor. The feedstock was exposed to high temperatures within the fixed bed reactor in the absence of oxygen. The flow of the nitrogen (N₂) carrier gas created an inert atmosphere within the system. As the feedstock was heated, it experienced physical and chemical separation into different molecules. The thermal devolatilization led to the formation of products in the form of solids (char) and volatiles (condensable and non-condensable or permanent gases) (Hirvonen, 2017). The liquid only formed from the condensable volatiles when it cooled in the condenser to allow separation into pyrolysis oil and permanent gas. The cooling system used as a part of the pyrolysis unit was a chiller and an ice bath which was connected to the condenser to enable a cold temperature ranging between 1-3°C. Three collectors ensued after the condenser, and these were placed in dry ice to ensure sufficient time to allow further condensation of the high volatile condensables to liquid products from the leather feedstock. The inclusion of the last two condensers was motivated by the additional liquid fractions observed in these condensers. These condensers allowed for further condensation and therefore accounted for all liquid fractions. The non-condensable gases flowed through a vent into an extractor fan.

2.3 Experimental design

The processing factors considered in the experimental design of this study were temperature, solid residence time, and N_2 flow rate. The temperature range, solid residence time, and center points were chosen based on the typical ranges cited in the literature and the operability range of the existing pyrolysis setup.

Operating temperature is the most critical parameter in Thermochemical Conversions (TCCs) because the temperature of the reactor is directly correlated to the product yields obtained (He et al., 2000). For this study, the operating temperatures chosen were 450, 550, and 650°C, including lower and higher axial points of 409 and 691°C, respectively (Table 3). Previous studies were considered when selecting the temperature range used in the present study (Yilmaz et al., 2007, Marcilla et al., 2012, and Kluska et al., 2019). The center point temperature chosen for pyrolysis (550°C) was motivated by the fact that pyrolysis appears to be complete by this temperature as the majority of the sample devolatilization has taken place in this range. Choosing the temperature range of 450 and 650°C with a center point of 550°C enables the researcher to draw parallels with previous studies to verify whether pyrolysis of leather shoe waste is achievable in a comparable manner to leather tannery waste (Yilmaz et al., 2007, Marcilla et al., 2012, and Kluska et al., 2019). The use of different temperature ranges in the pyrolysis reactor enabled the researcher to determine the differences in product yields and the optimal conditions for leather footwear pyrolysis.

Solid residence time refers to the amount of time that the feedstock remains inside the reactor (Gao et al., 2017). The solid residence time used in pyrolysis studies is an important parameter that can influence product yields (Dhyani and Bhaskar, 2018). For this study, the solid residence time reflects the amount of time the sample was held inside the reactor once the processing temperature was met. The hold times used in this study had a center point of 10 minutes and lower and upper levels of 5 and 15 minutes, respectively (Table 3). The lower and higher axial points are 3 and 17 minutes, respectively. The continuous flow of N2 gas flow created an oxygen-free atmosphere within the reactor (Aziz et al., 2018). An inert atmosphere is imperative as it creates conditions which are chemically inactive to ensure pyrolysis takes place without the risk of contamination from oxidizing gases such as oxygen (Jo et

TABLE 3: Central composite design (CCD) factors.

Factor	Unit	Factor type	Lower level (-1)	Center point (0)	Upper level (+1)	Lower axial (-1.414)	Higher axial (+1.414)
Temperature	°C	Variable	450	550	650	409	691
Solid residence time	min	Variable	5	10	15	3	17
Nitrogen flow rate	L/min	Constant	±6.5	±6.5	±6.5	±6.5	±6.5
Particle size	cm	Constant	0.5	0.5	0.5	0.5	0.5
Sample amount	g	Constant	40	40	40	40	40
Solid yield	wt.%	Response					
Liquid yield	wt.%	Response					
Gas yield	wt.%	Response					

al., 2017). The nitrogen flow rate (99.5% purity, Afrox, South Africa) remained constant at 6.5L/min for the pyrolysis experiments in this study. It was essential to ensure the nitrogen flow rate remained constant to provide a controlled inert atmosphere and to prevent counter airflow that may have compromised the experimental runs.

This study used a two-level full factorial design with a replication of experiments to verify the results. A central composite design (CCD) was employed with three center points, and an overview of the design and response factors are provided in Table 3. This experimental design enabled the researcher to investigate the optimization of operating conditions for pyrolysis yields. A statistical analysis of the variance was also employed using Minitab® 19 Statistical Software. A response surface model was fitted to determine the effects of temperature and solid residence time on each of the pyrolysis yields. Moreover, this also aided the researcher in identifying conditions that optimized the pyrolysis yields within the study.

2.4 Mass balance

The mass balance was a central component used to calculate the yields for each experimental pyrolysis run. The reactor and condensers were weighed before and after each pyrolysis experiment to calculate the percentage of solid, liquid, and gas yields. The difference in weight within the reactor was attributed to the remaining char whilst the difference in the condenser weight was attributed to the liquid yield. The gas yield was calculated using the difference in weight, and hence the remaining fraction was allocated to the gas yield.

2.5 Analytical techniques

2.5.1 Thermogravimetric analysis

Thermogravimetric analysis (TGA) was one of the analytical methods employed in this study. Through TGA, it was possible to assess the weight loss function of temperature under the nitrogen atmosphere (Sethuraman et al., 2013). According to Januszewicz et al. (2016), TGA enables users to determine the characteristics of the temperature dependence of weight loss at a constant heating rate. TGA assumes that several products can form from a single substrate (Januszewicz et al., 2016). It can, therefore, be inferred that the number and types of products formed from the leather waste were proportional to changes in mass decomposition.

In this study, TGA experiments were performed on chrome-tanned leather waste using a sample mass of 10 ± 0.5 mg and particle size ranges of 0.5 to 0.8mm. The samples were inserted into 70 µL alumina sample holders and experiments were performed using the DTG-60AH detector (Shimadzu, Kyoto, Japan). The temperature range was between 25 and 800°C, and the heating rate was 25°C/min. An inert atmosphere was created using nitrogen (99.5% purity, Afrox, South Africa) with a flow rate of 80ml/min. TGA experiments were carried out three times to ensure reproducibility of the observed results.

2.5.2 Elemental analysis

The second analytical technique used in this study was an elemental analysis, which is also known as an ultimate analysis. This is the process through which the sample of leather material was analyzed for its elemental composition (Basu, 2018). The elemental composition in terms of Carbon, Hydrogen, Nitrogen, and Sulfur (CHNS) was determined for the raw leather shoe waste, the residual char, and the liquid oil fractions. This analysis was used to determine the ratio of elements from within the leather samples and the composition of the solid and liquid pyrolytic yields. The instrument used for this analysis was the Thermo Fisher Scientific Flash 2000 CHNS-O Organic Elemental Analyzer. The results of the ultimate analysis (C, H, N and O content) were also used to estimate the higher heating values (HHVs) of the pyrolysis product fractions using the correlation equation (1) (Mkhize et al., 2015, and Demirbas, 2016). The HHV is significant as it provides an indication of the amount of heat released and hence the energy density of the solid and liquid fractions.

$$HHV = 33.5[C] + 142.3[H] - 15.4[O] - 14.5[N] \times 10^{-2}$$
(1)

2.5.3 Gas chromatography-mass spectrometry

Gas chromatography-mass spectrometry (GC-MS) (Shimadzu GC-MS QP2010SE series model) was used to detect compounds from the liquid yield. The compounds were detected using the gas chromatographic retention times and identified in the mass spectrometer. The sample was swept through a capillary column (Zebron ZB-5MSplus 30m x 0,25mm, 0,25um id). using helium (99.9% purity, Air products, South Africa) as the carrier gas and components were separated based on the volatility and affinity of the

mixture. The method used for the analysis was as follows: the initial temperature was set at 40°C; the final temperature was set at 270°C; the heating rate was 10°C per minute, and the total time for the run was 48 minutes. A qualitative analysis of the chromatogram was conducted using the NIST mass spectral library (version 4.0) to identify the compounds present in the liquid yield. Since the qualitative analysis was used the researcher emphasized determining what constituted the liquid yields rather than determining exactly how much of each compound was present. The qualitative nature of the study is elaborated on in the following section.

2.6 Concept proving and desirability study

This research was premised primarily as a concept proving and desirability study. The focus was placed on establishing whether chrome tanned leather from post-consumer footwear has the potential to be pyrolyzed. Pyrolysis experimental runs were used to determine optimal conditions for pyrolysis product yields, and this constituted the desirability aspect of the study. The quantities of the pyrolysis yields were considered concerning the processing temperature and solid residence time. In addition, the emphasis was placed on the replication of pyrolysis experimental runs to ensure the validity of the research findings. The GC-MS had a qualitative focus in its methodology for the analysis of the liquid yields. The constituents of the liquid yields were analyzed in this study with a greater focus on the composition of the yield rather than the quantities.

3. RESULTS AND DISCUSSION

3.1 Thermogravimetric analysis

The thermogravimetric analysis (TGA) of chrometanned leather samples for a heating rate of 25°C/min revealed that the first mass loss of the sample is apparent between 0 and 150°C (Figure 2). The mass loss observed in this temperature range is attributed to the release of inherent moisture contained within the sample. Yilmaz et al. (2007) describe this component of the liquid yield as the aqueous phase. An explanation for this peak is that the pyrolysis temperature reached close to 100°C, which is the temperature at which water evaporates (Kluska et al., 2019). According to Pham et al. (2018) water absorbs heat and uses it to evaporate and can also play a role in delaying devolatilization. The majority of the mass loss is observed between 250 and 550°C, with the most significant mass release peak occurring at 350°C (Figure 2). A 50% reduction in mass was experienced between 250 and 550°C resulting in the large weight loss peak in the DTG curve. This mass release is caused by the thermal devolitilisation of volatile organic compounds present in the leather sample. The maximum temperature peak demonstrates the significant impact that heat transport has on forming the mass loss curves. The TGA curve begins to flatten after 550°C indicating the near completion of pyrolysis. Additionally, it is observed that after pyrolysis is complete, the remaining mass accounts for approximately 25 wt.% of the sample (Figure 2).

3.2 Pyrolysis yields from leather shoe waste

This section outlines the product yields as a result of pyrolyzing leather shoe waste. Available data from the literature indicates that the mass balance of different types of pyrolysis products is dependent on the final temperature in the pyrolysis process (Yilmaz et al., 2007, and Xiao and Yang, 2013). The effect of temperature and solid residence time on the pyrolysis yields (solid, liquid, and gas) were investigated through CCD experiments according to the parameters outlined in Table 3. The results obtained from the CCD experiments are summarized in Table 4. The averages of the solid, liquid, and gas yields are presented in conjunction with the standard deviation (SD) in Table 4.

3.2.1 Solid yield

The solid yield consisted of char, a carbonaceous material generated through the thermal devolatilization of leather in the pyrolysis reactor. The solid yields ranged between



FIGURE 2: TGA and mass loss curve for leather waste in a nitrogen atmosphere with a heating rate of 25°C/min.

TABLE 4: Central Composite Design (CCD) of experiments.

Run no.	Pyrolysis pa	rameters	Leather pyrolysis products				
	Temperature (°C)	Time (min)	Solid yield (wt.%)	Liquid yield (wt.%)	Gas yield¤ (wt.%)		
			Average ±SD	Average ±SD	Average ±SD		
1.	409	10	42.63±0.88	38.43±3.01	19.00±2.12		
2.	450	5	38.38±2.30	45.50±1.06	16.13±3.36		
3.	450	15	33.50±0.71	46.88±2.30	21.30±5.13		
4.	550	3	29.63±1.59	49.63±3.36	20.75±4.95		
5.	550	10	27.75±2.47	52.63±1.94	19.63±4.42		
6.	550	17	27.75±2.30	40.38±1.94	32.25±4.24		
7.	650	5	24.75±1.06	51.75±2.47	23.50±3.54		
8.	650	15	24.63±0.88	46.25±2.12	29.13±3.01		
9.	691	10	24.50±0.00	46.63±2.30	28.88±2.30		

^a By difference

42.63 to 35.94 wt.% at temperatures of 409 and 450°C, respectively (Table 4). This change in mass indicates that at lower temperatures, pyrolysis was incomplete. Molecular components were unable to devolatilise at lower temperatures fully and only the weak chemical bonds were broken (He et al., 2018). These yields are congruent with previous studies that revealed solid fractions from leather tannery waste between a similar range of 33 and 38.5 wt.% at 450°C (Marcilla et al., 2015, Kluska et al., 2018). Moreover, at 550°C, the present study indicated a mass of 28.25 wt.% that is consistent with the yields obtained by Marcilla et al. (2012). The similar solid yields between studies reveal that leather waste from shoes can be pyrolyzed to yield similar results to leather tannery waste.

The response surface model in Figure 3 provides a visual representation of the interactions between temperature and hold time on the solid yield. It is observed that there is a negative correlation between temperature and solid yield because an increase in temperature corresponds to a decrease in the solid yield. As the temperature increases past 600°C, the response surface begins to

flatten with lower yields indicating the near completion of pyrolysis. The relationship between solid residence time and solid yield appears to be less significant compared to temperature. This is because only minor differences in the solid fraction distribution were observed as a function of the hold time.

The solid yield reached a constant value of approximately 25 wt.% at 650°C, indicating the completion of leather pyrolysis under CCD conditions. The completion of pyrolysis can be verified since the solid yield remaining corresponds to the fixed carbon and ash proportions of the leather sample in Table 2. This is further verified through an examination of the remaining mass in TGA experiments which accounts for approximately 25 wt.% of the sample (Figure 2). The char yields obtained through pyrolysis operating at two different scales are illustrated in Table 5. The fixed bed pyrolyzer operated using a gram-scale whilst the TGA operated using a microgram-scale. The yields are comparable since the solid fractions are similar for both setups. The slight variations in the wt. % may be attributed



FIGURE 3: . Response surface model illustrating the effect of temperature and hold time on solid yield.

TABLE 5: Solid yields	for CCD pyrolysis	and TGA experiments
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Temperature (°C)	Fixed bed pyrolyzer (wt. %)	TGA (wt. %)
409	42.63	46.85
450	36.94	39.17
550	28.25	30.45
650	24.69	28.45
690	24.5	27.56

 TABLE 6: Elemental composition of char obtained from leather shoe waste.

Components	Char (wt. %) ^a	TGA (wt. %)
Nitrogen	8.69	46.85
Carbon	62.67	39.17
Hydrogen	2.36	30.45
Sulfur	0.74	28.45
Oxygen ^b	12.98	27.56

to differences in the heating rates across the two setups. TGA had a controlled heating rate whilst the fixed bed pyrolyzer had a heating rate that was more variable and hence less constant in comparison. The comparability of the findings at different scales, as well as the consistency of the char yields with the fixed carbon and ash content, verifies the completion of pyrolysis under CCD conditions and further suggests the suitability of leather shoes for pyrolysis technology.

The elemental composition of char generated from leather shoe waste pyrolysis is shown in Table 6, with carbon taking the largest share of the composition. The higher heating value of the solid fraction was determined to be 25.55 MJ/kg. The char generated through pyrolysis could be used in the metallurgical sector. According to Filho et al. (2016), carbonized leather residues have the potential to partially replace coal in the metallurgical process, especially if they have a high heating value of 20.8MJ/kg. Since the HHV was 25.55 MJ/kg in this study this reinforces its potential use in this sector. Moreover, Bañón et al. (2016) pointed out that char may be used as cheap fuel, either pure or blended with other fuels. Yilmaz et al. (2007) also reported that char products from pyrolyzed leather could be used in the production of activated carbon and as an adsorbent of dyes from aqueous solutions.

3.2.2 Liquid yield and analysis

The liquid yield ranged between 36.25 and 54 wt.% at temperatures of 409 and 550°C, respectively. An increase in the process temperature corresponded to a rise in the liquid yield up to 550°C. It has been well documented that tar formation rate should increase with an increase in the pyrolysis temperature (Xiao and Yang, 2013, Bañón et al., 2016, and Baniasadi et al., 2016). The liquid fraction was maximized within the temperature range of 550-650°C. This finding is affirmed in Figure 4 since the liquid yield peaks between 550 and 650°C. The convex profile of the response surface reveals a well-defined optimum condition for the liquid yield. The highest liquid yield was achieved at 550°C with a hold time of 10 minutes (54 wt.%), and a close second was observed at 650°C with a hold time of 5 minutes (53.5 wt.%) (Table 4). The current findings are supported by Marcilla et al. (2012) and Kluska et al. (2019), who observed the maximum liquid yields within a similar temperature range of 500-550°C. The general findings from the CCD experiments and the convex shape of the response surface suggest that leather waste from shoes can be pyrolyzed between an optimal temperature range of 550-650°C.

Lower liquid yields were evident at lower temperatures corresponding with high solid yields. For instance, a liquid yield of 36.25 (wt.%) was reached at 409°C whilst solid yields remained high, accounting for 42.63 (wt.%) of the yield at 409°C. Since polymeric components could not fully



FIGURE 4: Response surface model illustrating the effect of temperature and hold time on liquid yield.

devolatilise at lower temperatures, this explains the high char yields at lower temperatures and corresponding low liquid yields.

The liquid fraction comprised of the aqueous phase (water content) and the organic phase (oil) (Yilmaz et al., 2007). The approximate weight basis ratio of the liquid content is 1-part oil to 3-parts water (1:3). The water-soluble fraction consists of lower molecular weight substances whilst the oil consists predominantly of brown tar that contains higher molecular weight compounds. The elemental composition of the oils obtained from leather shoe waste is shown in Table 7. It should be noted that the elemental weighting is comparable to those reported from chrome-tanned leather shavings in the study by Yilmaz et al. (2007). In both studies, carbon had the highest share of the elemental composition.

The high presence of carbon from the ultimate analysis indicates a high energy density within the leather pyrolysis oil. The higher heating value (HHVs) of the liquid fraction was determined to be 33.62MJ/kg at 550°C. This value is significantly higher than HHVs for many other waste pyrolysis oils. For instance, Table 8 shows a comparison of the HHVs between leather oil, biomass derived pyrolysis oils and conventional oils. The HHV of the leather pyrolysis oils are higher than biomass pyrolysis oils such as softwood lignin (30.04 MJ/kg) (Ben and Ragauskas, 2011), softwood barks (25.3-26.7 MJ/kg)(Ben et al., 2019), wood and agricultural residues (16.0-19.0 MJ/kg)(Zhang et al., 2007, and Meier et al., 2013) and pine/spruce wood (16.4-17.6 MJ/kg) (Chiaramonti et al., 2007). In addition, the leather oil HHVs are higher than low-grade coal (18 MJ/kg) (Dincer and Zamfirescu, 2014) but lower than conventional oils such as diesel (45.7 MJ/kg) and gasoline (47.3 MJ/kg) (Channiwala and Parikh, 2002). The HHV of the leather oil therefore indicates it has potential as an energy source or fuel.

3.2.3 Gas yield

In general, the gas fraction comprised a smaller share of the product yield in comparison to the solid and liquid fractions. The results indicate that, within the evaluated temperature range, the highest gas yield occurred at 550°C and 650°C during experimental runs that had a longer hold time. The lowest gas yield was observed at 450°C with a short hold time of 5 minutes. The low gas yield is likely attributed to the incomplete pyrolysis at this temperature, which accounts for the corresponding high solid yield. Higher process temperatures led to an increase in the gas yields at 550°C and 650°C whilst the lower process tem-

TABLE 7: Elemental	composition	of oil	obtained	from	leather	shoe
waste.						

Components	Oil (wt. %)
Nitrogen	8.33
Carbon	69.64
Hydrogen	8.57
Sulfur	1.16
Oxygen ^b	12.98
^b By difference.	

 TABLE 8: Comparison of HHVs for leather pyrolysis oil, other waste pyrolysis oils and conventional fuels.

Fuel type	HHVs (MJ/kg)	Reference
Leather oil	33.6	Present study
Biomass derived pyrolysis o	ils	
Softwood craft lignin	30.04	(Ben and Ragauskas, 2011)
Softwood barks (Raw pine, Douglas-Fir and Pine)	25.3- 26.7	(Ben et al., 2019)
Wood and agricultural residues	16.0- 19.0	(Zhang et al., 2007)
Wood-derived bio-oil	17.0	(Meier et al., 2013)
Pine/spruce wood	16.4- 17.6	(Chiaramonti et al., 2007)
Low-grade coal	18	(Dinçer and Zamfirescu, 2014)
Conventional oils		
Diesel	45.7	(Channiwala and Parikh, 2002)
Gasoline	47.3	(Channiwala and Parikh, 2002)

perature of 450°C with a hold time of 5 minutes caused the lowest gas yield. This finding was consistent with the work of Yilmaz et al. (2007), which found similar trends of increasing gas yields and decreasing char yields as a function of higher temperatures.

A visual representation of the gas yield distribution as a function of temperature and hold time is shown in Figure 5. The increase in the gas yield can be correlated to factors including the fragmentation of the longer chained condensable volatile molecules and the sublimation of the char to permanent gases. Holding time had a less significant influence on gas yield in comparison to processing temperature. A possible explanation for this is because gas flow, heating rate and residence time of the hot condensable volatiles have a greater influence on the fragmentation of heavy molecules to light molecules in comparison to holding time. In addition, another influential factor was the fairly rapid reaction during pyrolysis which meant that even at lower hold times pyrolysis was near completion, which influenced the gas yield distribution. The response surface in Figure 5 reveals that the correlation between variables and the gas yield is less significant in comparison to trends found in the solid and liquid yields response surface. The variability in the gas yield correlating to an increase in temperature and hold time is supported by results found by Yilmaz et al. (2007). In the study by Yilmaz, a final temperature increase from 450 to 600°C caused an increase in gas yield from 17 to 23 wt.% similar to this study. Similarly, Marcilla et al. (2012) observed that gas yields tended to rise with an increase in process temperature.

3.3 Gas chromatography-mass spectrometry analysis

The results of the GC-MS analysis of leather shoe waste are presented in this section. Each of the liquid fractions from the CCD pyrolysis experiments was analyzed using the GC-MS. In total, nine liquid yields were analyzed with an initial and repeated run to ensure the replicability of findings. The general trends are discussed to provide a concise synopsis of the types of compounds present in the liquid fraction.

The results of the GC-MS analysis show a complex spectrum of liquid products from leather shoe waste. The ten major compounds identified in each of the experiments are summarized in Table 9. The main compounds identified in each of the experiments were italicized in Table 9 to emphasize their significance. The majority of the detected compounds correspond to nitrogen derivatives, which is related to the protein content of the leather shoe waste (Font et al., 1999, Sethuraman et al., 2014, and Kluska et al., 2019). The major compounds identified across GC-MS runs are 9-Octadecenamide, (Z)-; Octadecanamide, 9-Octadecenoic acid, 1,2,3-propanetriyl ester; n-Heptadecanol-1 and 13-Docosenamide. It is noteworthy that 9-Octadecenamide, (Z)-; 9-Octadecenoic acid, 1,2,3-propanetriyl ester; and 13-Docosenamide are isomers since there is a different positioning of the functional group at position 9 and 13, respectively, instead of the normal position. It is important to take note of isomers given the differences in the arrangement of atoms within the molecule, despite the similarity in the molecular formula.

On average, the chromatograms of the liquid yield identified between 100-150 different peaks. The spectrum of compounds formed were complex, most of which were attributed to protein decomposition. As an example, Table

10 shows the list of compounds identified by the GC-MS analyzer from pyrolytic liquid yield at 550°C with a hold time of 10 minutes. The detected compounds in the liguid yield correspond to the chromatogram peaks depicted in Figure 6. Each peak in the chromatogram represents a compound present in the sample. The x-axis of the chromatogram represents the retention time(min), and the y-axis represents the detector response and hence the peaks of the different compounds. The motivation for presenting the results of only one of the chromatograms and tables is because there was minimal discrimination between the composition of the liquid sample yields, and a summary of the significant compounds is depicted in Table 9. The similarity index (SI) values for each peak are also presented in Table 10, according to the mass spectra libraries used. SI compares the query mass spectra with the reference mass spectra via spectrum matching and hence indicates the degree of certainty of the identified compound (Wei et al., 2014). Approximately 41% of the peaks in Table 10 present SI values greater than 90%. The relatively high SI values, therefore, suggest that there was a high level of certainty amongst the identified compounds.

The main types of compounds identified in the sample were grouped based on their structure. Table 11 depicts the percentage of compounds classified according to their composition. The main group of compounds present in

TABLE 9: GC-MS chromatogram main compound list for the different pyrolytic liquid fractions.

Pyrolytic liquid fractions	Compound name	Retention time (min)
	Oleanitrile	30.42
	9-Octadecenoic acid, 1,2,3-propanetriyl ester	31.24
	Octadecanamide	31.51
	N-Methyldodecanamide	31.71
Viold 1 (AEO°C: Emin)	Cyclobutane, 3-hexyl-1,1,2-trimethyl-, cis-	31.84
field I (450 C, Sifilit)	Triethylene glycol monododecyl ether	32.11
	9-Octadecenamide, (Z)-	33.04
	Octadecanamide	33.23
	9-Octadecenamide, N,N-dimethyl-	33.55
	Pyrrolidine, 1-(6-methyl-1-oxooctadecyl)-	35.08
	Phosphonic acid, (p-hydroxyphenyl)-	16.32
	Caprolactam	21.28
	5,10-Diethoxy-2,3,7,8-tetrahydro-1H,6H-dipyrrolo	29.31
	n-Heptadecanol-1	30.42
Viold 2 (6E0°C) E min)	Hexadecanamide	31.44
field 2 (650 C, 5 min)	Coumarin-3-carbonitrile, 6-methyl-	31.66
	1,8-Diazacyclotetradecane-2,9-dione	32.64
	9-Octadecenamide, (Z)-	33.03
	9-Octadecenamide, (Z)-	33.08
	Octadecanamide	33.23
	(1H)Pyrrole-2-carbonitrile, 5-methyl-	20.98
	Phenol, 2-undecyl-	21.06
	5,10-Diethoxy-2,3,7,8-tetrahydro-1H,6H-dipyrrolo	29.24
	n-Heptadecanol-1	30.42
Viold 2 (450°C: 15min)	Octadecanamide	31.43
	Phenol, 4,4'-(1-methylethylidene)bis-	31.75
	Benzeneacetic acid, 4-pentadecyl ester	31.87
	9-Octadecenamide, (Z)-	33.01
	9-Octadecenamide, (Z)-	33.06
	Octadecanamide	33.20

	Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-	27.64
	5,10-Diethoxy-2,3,7,8-tetrahydro-1H,6H-dipyrrolo	29.25
	Oleanitrile	30.44
	6-Octadecenoic acid, (Z)-	31.31
	Octadecanamide	31.54
Yield 4 (650°C; 15min)	9-Octadecenamide. (Z)-	33.09
	13-Docosenamide. (Z)-	33.14
	Octadecanamide	33.28
	N-Mothyldodooanamido	22.25
		22 57
		15.02
	Annine	15.22
		21.09
		28.38
	5,10-Diethoxy-2,3,7,8-tetrahydro-1H,6H-dipyr	29.25
Yield 5 (550°C; 10min)	n-Heptadecanol-1	30.40
	Octadecanamide	31.44
	Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(31.87
	Triethylene glycol monododecyl ether	32.09
	9-Octadecenamide, (Z)-	33.05
	Octadecanamide	33.19
	Caprolactam	20.92
	5,10-Diethoxy-2,3,7,8-tetrahydro-1H,6H-dipyrrolo	29.20
	1-Hexadecanol	30.41
	9-Octadecenoic acid, 1,2,3-propanetriyl ester	31.26
Viold $(400^{\circ} \text{C}; 10 \text{ min})$	Octadecanamide	31.53
field 6 (409 C, 10 min)	9-Octadecenamide, (Z)-	33.04
	Octadecanamide	33.23
	9-Octadecenamide, N,N-dimethyl-	33.55
	9-Octadecenoic acid (Z)-, 2-butoxyethyl ester	35.41
	Cholesta-3,5-diene	40.02
	5,10-Diethoxy-2,3,7,8-tetrahydro-1H,6H-dipyrrolo	29.28
	Oleanitrile	30.45
	Hexadecanenitrile	30.62
	6-Octadecenoic acid. (Z)-	31.40
	Octadecanamide	31 63
Yield 7 (691°C; 10 min)	N-Methyldodecanamide	31.69
	1H-Isoindole-13(2H)-dione 2-(2-propynyl)-	31 77
	13-Docosenamide (7)-	33.16
	Octodoconomido	22.22
		22 60
		33.36
	Oreanitine	30.39
		30.82
		31.01
	6-Octadecenoic acid, (Z)-	31.33
Yield 8 (550°C; 17min)	Octadecanamide	31.56
	N-Methyldodecanamide	31.64
	D-Mannohexadecane-1,2,3,4,5-pentaol	31.70
	Cyclopropanecarboxylic acid,pentadecyl ester	32.03
	9-Octadecenamide, (Z)-	32.87
	13-Docosenamide, (Z)-	33.03
	Phenol, 2-methyl-	17.33
	1H-Indene, 1-methylene-	19.56
	Caprolactam	21.08
	Naphthalene, 2-methyl-	21.57
Vield Q (550°C; 2min)	5,10-Diethoxy-2,3,7,8-tetrahydro-1H,6H-dipyrrolo	29.20
11eiu 9 (330 C, 311111)	n-Heptadecanol-1	30.34
	Octadecanamide	31.37
	9-Octadecenamide, (Z)-	32.93
	9-Octadecenamide, (Z)-	32.98
	Octadecanamide	33.21
	•	•

Peak #	R Time	Area %	Formula	A/H Name	SI (%)
1	11.767	0.20	C6H11N	Hexanenitrile	95
2	13.692	0.21	C6H9N	1H-Pyrrole, 2,5-dimethyl-	92
3	15.224	1.79	C6H7N	Aniline	96
4 15.380 0.64 C7H5N		C7H5N	Benzonitrile	95	
5	15.870	0.61	C6H704P	Phosphonic acid, (p-hydroxyphenyl)-	96
6	15.932	1.62	C6H6O	Phenol	96
7	16.000	0.34	C7H11N	1H-Pyrrole, 2-ethyl-4-methyl-	88
8	16.181	0.50	C5H6N2	2-Aminopyridine	95
9	16 458	0.92	C10H16	D-Limonene	95
10	16 560	0.22	C8H180	1-Hevanol 2-ethyl-	88
10	16.000	0.22	Соносі	1H-Indene 1-chloro-2 2-dihvdro-	05
10	17 260	1 15	0711901	Phonol 2-mothyl-	07
12	17.309	0.07	C71100		97
13	17.413	0.27	C/H9N	p-Aminotoluene	92
14	17.476	0.40	C/H9N	p-Aminotoluene	98
15	17.856	1.04	C7H80	p-Cresol	95
16	17.935	0.51	C6H8N2	2-Pyridinamine, 3-methyl-	90
17	18.067	0.36	C8H7N	Benzonitrile, 4-methyl-	96
18	18.240	0.24	C8H100	Phenol, 2,5-dimethyl-	92
19	18.560	0.17	C7H120	1-Heptyn-4-ol	74
20	18.689	0.24	C8H18O2	1-Butanol, 4-butoxy-	92
21	18.725	0.18	C8H7N	Benzyl nitrile	92
22	19.095	0.32	C8H100	Phenol, 2,4-dimethyl-	89
23	19.521	0.71	C7H10N2	Pyridine, 2-aminomethyl-6-methyl-	70
24	19.581	0.53	C10H8	Naphthalene	98
25	19.695	0.43	C9H17N0	Octane, 1-isocyanato-	78
26	20.414	0.33	C9H9N	Benzenepropanenitrile	96
27	20.459	0.18	C8H10Cl2	Cyclohexene, 1-chloro-4-(1-chloroethenyl)-	88
28	20.684	0.32	C9H12O	Phenol. 3-(1-methylethyl)-	92
29	21.088	7.76	C6H11N0	Caprolactam	94
30	21 362	0.26	C11H10	Nanhthalene 2-methyl-	90
20 21	21 604	0.65	C11H10	Nanhthalene 2-methyl-	90 Q1
32	27.004	0.00	C5H8N2O2	2 /-Imidezolidinedione 5 5-dimethyl-	60
22	22.017	0.72	C0U16O2	Piovolo[2 2 1]nonono-2 0-diol ovo-anti-	67
24	22.040	0.20			0/
25	22.701	0.12	0120260		04
30	23.092	0.30	0120200	(havehidre 211 azenin 2 anete N1 02)his(1 methidethul) (t.4)	70
30	23.811	0.15			73
37	25.055	0.20	C5H4N4U	I H-Imidazo(4,5-d)pyridazin-7-oi	/8
38	25.400	0.37	C8H1802		80
39	25.497	0.12	C8H/NS2	Benzothiazole, 2-(methylthio)-	85
40	26.160	0.44	C16H30Cl202	Dichloroacetic acid, tetradecyl ester	81
41	26.603	0.18	C14H30O2	Ethanol, 2-(dodecyloxy)-	91
42	26.694	1.28	C11H10N20	Furaldehyde phenylhydrazone	75
43	26.894	0.95	C10H14N2O3	3-Methyl-1,4-diazabicyclo[4.3.0]nonan-2,5-dione, N-acetyl-	84
44	26.965	0.34	C10H140	Benzene, (1-methylpropoxy)-	64
45	27.110	0.12	C10H8N20	3-Phenoxypyridazine	77
46	27.210	0.53	C10H14N2O3	3-Methyl-1,4-diazabicyclo[4.3.0]nonan-2,5-dione, N-acetyl-	72
47	27.403	0.32	C17H31F3O2	Trifluoroacetic acid, pentadecyl ester	91
48	27.470	0.47	C14H29Br	2-Bromotetradecane	77
49	27.560	1.07	C11H240	Undecanol-4	77
50	27.620	1.36	C7H10N202	Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-	87
51	27.780	0.20	C15H22N2O4	2,6-Piperidinedione, 1,1'-(1,5-pentanediyl)bis-	73
52	28.181	0.11	C16H22O4	1,2-Benzenedicarboxylic acid, bis(2-methylpropyl) ester	89
53	28.377	2.42	C18H38O	1-Octadecanol	87
54	28.572	0.31	C15H29N	Pentadecanenitrile	94
55	28.941	0.47	C8H12O3	6-Hydroxy-9-oxa-bicyclo[3,3,1]nonan-3-one	69
56	29 145	0.41	C11H18N2O2	Pyrrolo[1.2-a]pyrazine-1.4-dione hexahydro-3-(2-methylnronyl)-	75
57	29 246	3 37	C14H22N2O2	5 10-Diethoxy-2 3 7 8-tetrabydro-1H 6H-dinyrrolo[1 2-a:1'2'-d]nyrazine	02 , 0
57	27.270	0.07	01112213202		20

59 29.517 0.20 C16H3403 Diethylene glycal monododcyl ether 99 60 30.105 0.28 C11H3002 2-Propanone, 1-(1-cyclohexen-1yl)-3-ethoxy- 59 61 30.174 0.21 C18H360 9-Octadecen-1-0(2)- 68 63 0.505 0.25 C17H3202 7-Hexadecenoic add, methyl eter. (2)- 73 64 30.57 0.26 C14H27N Tetradecenoic add, methyl eter. (2)- 73 64 30.57 0.26 C14H27N Tetradecenentrile 85 65 30.77 0.70 C7H9N50 C14H27N Benzemanne, 4/-methylenbia- 88 68 31.005 0.63 C18H38S 1-Octadecanethiol 87 70 31.43 1.53 C18H38N Octadecanamide 90 71 31.647 1.14 C18H37NO Octadecanamide 81 72 31.489 2.71 C11H18N202 Phynolol_2-alprazine 1.4-dione, hexahydro 3(2-methylentylopyl-)- 62 73 31.869 2.	58	29.346	1.26	C11H18N2O2	Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(2-methylpropyl)-	86
60 30.105 0.28 C11H1802 2-Propanon.1(-ryclohexen-1y)3-sethoxy. 59 61 30.174 0.21 C18H360 9-Octadecen1-0, (Z) 96 62 30.507 0.25 C17H300 1-Heptadecenol: acid, methyl ester, (Z). 73 63 30.507 0.70 C1H9X00 1-Heptadecenol: acid, methyl ester, (Z). 73 64 30.595 0.36 C1H427N Tetradeceneris acid, methyl ester, (Z). 78 65 30.677 0.70 C7H9N50 1-[3-Pyridyl]-3-amidine urea 65 66 30.733 0.12 C22H4002 Ethanol, Z-(sicosyloxy)- 89 67 31.015 1.53 C18H388 1-Octadecanethiol 77 71 31.447 1.14 C13H47N0 Octadecanamide 81 72 31.741 C17 C11H18N202 Pyrmolo[1-2-ajpyrazine-1.4-dione, hexapirdo-3 (2:methylprophyl)- 62 73 31.69 2.71 C11H18N202 Pyrmolo[1-2-ajpyrazine-1.4-dione, hexapirdo-3 (2:methylprophylprot) 62 <t< td=""><td>59</td><td>29.517</td><td>0.20</td><td>C16H34O3</td><td>Diethylene glycol monododecyl ether</td><td>90</td></t<>	59	29.517	0.20	C16H34O3	Diethylene glycol monododecyl ether	90
61 30.174 0.21 C181800 9-Octadecen1-ol, (2)- 96 62 30.403 3.16 C17H300 n-Heptadecanol-1 92 63 30.507 0.28 C17H3202 7.Hexadecenoic acid, methyl ester, (2)- 73 64 30.573 0.12 C17H3202 7.Hexadecaneitrile 85 65 30.677 0.70 C7H9N50 1-[3-Pyridyl]-3-amidine urea 65 66 30.793 0.12 C22H4602 Ethnol, 2/e(cosyloxy)- 89 67 31.011 0.81 C7H9N50 1-0-tadecanethiol 77 63 31.059 6.63 C13H1AV Benzemanine, 4.4-methylenebias 88 69 31.145 1.53 C18H380 1-0-tadecanethiol 97 70 31.499 8.69 C18H37N0 Octadecanamide 90 71 31.623 2.14 C13H27N2 Pyrotol(1-2)apyrazine 1.4 dione, hexalydro-3 (cmbrigroup)h)- 62 73 31.869 2.71 C11H18N202 Pyrotol(1,2-a)	60	30.105	0.28	C11H1802	2-Propanone, 1-(1-cyclohexen-1-yl)-3-ethoxy-	59
62 30.403 3.16 C17H300 Methylester, (2) 73 64 30.557 0.70 C17H3202 77+Hzadecenoic acid, methyl ester, (2) 73 65 30.677 0.70 C7H9N50 Tetradecamentriale 65 66 30.79 0.71 C7H9N50 Ethyl 2-cyanocrotonate 65 67 31.011 0.81 C7H9N02 Ethyl 2-cyanocrotonate 75 68 31.055 0.63 C13H14N2 Benzenamine, 4,4" methylenelis- 88 69 31.145 1.53 C18H385 1-0-todecamethiol 77 70 31.439 8.69 C18H37N0 Octadecamethiol 81 71 31.647 1.14 C13H202 Pyrrolo[12-a]pyrazine1.4-dione, hexalydro-3(2-methylpropyl)- 62 73 31.89 2.71 C11H18V02 Pyrrolo[12-a]pyrazine1.4-dione, hexalydro-3(2-methylpropyl)- 62 74 32.265 1.33 C2H450 Cyclobraxe.1.4-didecyl- 76 74 32.255 1.25 C12H1	61	30.174	0.21	C18H360	9-Octadecen-1-ol, (Z)-	96
63 30.507 0.25 C11H202 7-Hexadecenoic acid, methyl ester, (2)- 73 64 30.595 0.36 C14H27N Tetradecamenitrilie 85 65 30.677 0.70 C7H9NSO 113-Prind(4)-Samidine urea 65 66 30.793 0.12 C22H4602 Ethanol.2-(eicosyloxy)- 89 67 31.011 0.81 C7H9NSO Ethyl 2-cyanocrotonate 75 68 31.095 0.63 C13H14V2 Benzenamine, A4-methylenebis- 88 69 31.445 1.83 C18H37NO Octadecanamide 81 71 31.647 1.14 C13H2NO Phrenol, 44-(1-methylethylidenebis- 75 73 31.869 2.71 C15H1602 Phyrolo[12-a)pyrazine-1.4-dione, hexahydro-3-(2-methylpropyl)- 62 74 32.093 2.14 C18H37NO Octadecanamide 81 75 32.265 1.33 C26H52 Cyclohexane, 1.4-didecyl- 76 76 32.405 1.11 C19H202	62	30.403	3.16	C17H360	n-Heptadecanol-1	92
64 30.595 0.36 C14H27N Tetradecanenitrile 85 65 30.677 0.70 C7H9N50 1-[3-Pyridy]-3-amiltine urea 65 66 30.73 0.12 C22H4602 Ethanol.2 (ceicosyloxy)- 89 67 31.011 0.81 C7H9N02 Ethyl 2-cyanocrotonate 75 68 31.095 0.63 C13H14N2 Berzzenamine, 4/-methylenebis- 88 69 31.145 1.53 C18H38S 1-Octadecanentiol 87 70 31.439 8.69 C18H7N0 Octadecanamide 90 71 31.647 1.14 C13H27NO Phronol, 4.4'(-methylethylitylicalex)bis- 75 73 31.869 2.71 C11H18N202 Pyrrolo[12-a]pyrazine-1.4-dione, hexahydro-3(2-methylpropyl)- 62 74 32.052 1.11 C19H2802 m-Toluic acid, undec-2-enyl ether 77 75 32.265 0.33 C22H4002 Ethanol.2 (-eicosyloxy)- 91 79 32.725 0.81 C18H1802 </td <td>63</td> <td>30.507</td> <td>0.25</td> <td>C17H32O2</td> <td>7-Hexadecenoic acid, methyl ester, (Z)-</td> <td>73</td>	63	30.507	0.25	C17H32O2	7-Hexadecenoic acid, methyl ester, (Z)-	73
65 30.677 0.70 C719NS0 113-Priptly13-samidine urea 65 66 30.793 10.12 C22H4602 Ethanol, 2(elcosyloxy) 89 67 31.01 0.81 C7H9NO2 Ethanol, 2(elcosyloxy) 89 68 31.095 0.63 C18H14N2 Benzenamine, 44-methylenebis- 88 69 31.145 1.53 C18H37NO Octadecanemide 90 71 31.647 1.14 C18H37NO Octadecanamide 90 73 31.69 2.71 C15H1602 Phrenol, 4, 4'(1-methylethylidene)bis- 75 73 31.680 2.71 C11H18N202 Pyrrolol, 2-gyrazine 1.4-dione, hexahydro-3 (2-methylpropyl)- 62 74 32.093 2.14 C18H3804 Triethylene glycol monododecyl ether 88 75 32.265 1.31 C26H52 Cyclohexane, 1,4-didecyl- 76 74 32.265 0.73 C22H4602 Ethanol, 2-(elcosyloxy)- 91 74 32.265 1.35 C12H18020	64	30.595	0.36	C14H27N	Tetradecanenitrile	85
66 30.793 0.12 C2214602 Ethand, 2-(eicosyloxy)- 99 67 31.011 0.81 C7H9N02 Ethyl 2-cyanocrtonate 75 68 31.059 0.63 C13H14N2 Benzenamine, 4-4'-methylenebis- 88 69 31.145 1.53 C18H38S 1-Octadecanethiol 87 70 31.439 8.69 C18H37N0 Octadecanamide 90 71 31.447 1.14 C13H27N0 N-Methyldodecanamide 81 72 31.741 2.07 C15H1602 Pyrolo[1,2-a]pyrazine1.4-dioce, hexatypro-3-2c methylsorpyl)- 62 73 31.669 2.71 C11H18N202 Pyrolo[1,2-a]pyrazine1.4-dioce, hexatyprolyces 75 74 32.025 1.33 C26H52 Oyclohexane, 1.4-didecyl- 76 75 32.625 0.73 C22H4602 Ethanol, 2-(eicosyloxy)- 91 77 32.525 1.25 C12H22N202 1.39 C14H3800 9-Octadecanamide 95 80 33.050	65	30.677	0.70	C7H9N50	1-[3-Pyridyl]-3-amidine urea	65
67 31.011 0.81 C7H9N02 Ethyl 2-cyanocrtonate 75 68 31.055 0.63 C13H14N2 Benzanamine, 4.4'-methylenebis- 88 69 31.143 1.55 C18H38X In-Octadecanarhide 900 71 31.647 1.14 C13H27N0 N-Methyldodecanarhide 81 72 31.741 2.07 C15H1602 Phronol, 4.4'(-Intethylethyldoreb)is- 75 73 31.869 2.71 C11H18N202 Pyrrolo[1,2-a]pyrazine 1.4-diodecyl ether 88 75 32.255 1.33 C26H52 Cyclohexane, 1.4-didecyl 76 74 32.050 1.11 C19H2802 m-Toluic acid, unde-2-englester 77 75 32.255 1.25 C12H4020 Ethanol, 2-(elcosyloxy)- 91 79 32.725 0.81 C18H37N0 Octadecanamide 95 80 33.050 11.86 C18H37N0 Octadecanamide 95 81 33.44 0.55 C21H32N0 Octadecanamide, (66	30.793	0.12	C22H46O2	Ethanol, 2-(eicosyloxy)-	89
68 31.095 0.63 C13H14N2 Benzenamine, 4/-inettylenebis- 88 69 31.145 1.53 C18H38S 1-Octadecanamide 90 71 31.647 1.14 C13H27N0 Octadecanamide 81 72 31.741 2.07 C15H1602 Phenol, 4.4 (1-mettylethylidene)bis- 75 73 31.869 2.71 C11H18N202 Pyrolo[1,2-a]pyrazinc-1,4-diney, hexahydro-3(2-methylpropyl)- 62 74 32.093 2.14 C18H3804 Triethylene glycol monododcyl ether 88 75 32.265 1.33 C26H52 Cyclohexane, 1.4 -dildeyl- 76 74 32.052 0.73 C12H2X02 1.8-Dizacyclotetradecane-2.9-dione 90 78 32.625 0.73 C22H4602 Ethanol, 2-(eicoylox)- 91 79 32.725 0.81 C18H1802 -4.4-1-dilyloxydipheryl 55 81 33.194 3.84 C18H37N0 Octadecanamide 95 82 33.370 1.17 C14H16N2	67	31.011	0.81	C7H9NO2	Ethyl 2-cyanocrotonate	75
69 31.145 1.53 C18H38S 1-Octadecanethiol 67 70 31.439 8.69 C18H37N0 Octadecanamide 90 71 31.447 1.14 C13H27N0 N-Methyldodecanamide 81 72 31.741 2.07 C15H1602 Phenol, 4.4'(1-methylethylidene)bis- 75 73 31.869 2.71 C11H18N202 Pyrrolo[1,2-alpyrazine-1,4-dicene, hz-Alpydo-3-(2-methylepropyl)- 62 74 32.093 2.14 C18H3804 Triethylene glycol monododecyl ether 88 75 32.265 1.33 C26H52 Cyclohexane, 1.4-didecyl- 76 76 32.405 1.11 C19H2802 m-Toluic acid, undec-2-enyl ester 77 78 32.625 0.73 C22H4602 Ethanol, 2-(eicosylow)- 91 79 32.725 0.81 C18H3N0 Octadecanamide (2)- 88 81 33.405 11.86 C18H3N0 Octadecanamide (2)- 88 82 33.370 1.17 C14H16N202 <td>68</td> <td>31.095</td> <td>0.63</td> <td>C13H14N2</td> <td>Benzenamine, 4,4'-methylenebis-</td> <td>88</td>	68	31.095	0.63	C13H14N2	Benzenamine, 4,4'-methylenebis-	88
70 31.439 8.69 C18H37N0 Octadecanamide 90 71 31.647 1.14 C13H27N0 N-Methyldotecanamide 81 72 31.741 2.07 C15H1602 Phron0, 4,4'1-methylethylidene)bis- 75 73 31.869 2.71 C11H18020 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(2-methylpropyl)- 62 74 32.093 2.14 C18H3804 Triethylene glycol monododecyl ether 88 75 32.265 1.33 C26H52 Cyclohexare, 1,4-didecyl- 76 77 32.525 1.25 C12H22N202 1.8-Diazacyclotetradecane-2,9-dione 90 78 32.625 0.73 C22H4602 Ethanol,2-(ejcasyloxy)- 91 79 32.725 0.81 C18H1802 4.4'-Diallyloxydiphenyl 55 80 33.050 11.86 C18H35N0 9-Octadecenamide, (2)- 88 81 33.194 3.84 C18H37N0 N-Methyldodecanamide 86 82.625 1.77 C214H6022	69	31.145	1.53	C18H38S	1-Octadecanethiol	87
71 31.647 1.14 C13H27N0 N-Methyldodecanamide 81 72 31.741 2.07 C15H1602 Phenol, 4.4'(1-methylethyllethyllethylethylethylethyleth	70	31.439	8.69	C18H37NO	Octadecanamide	90
72 31.741 2.07 C15H1602 Phenol, 4.4'(1-methylethylidene)bis- 75 73 31.869 2.71 C11H1BN202 Pyrrolo[1,2-a]pyrazine1,4-dione, hexahydro-3-(2-methylpropyl)- 62 74 32.093 2.14 C1BH3804 Triethylene glycol monododecyl ether 88 75 32.265 1.33 C26H52 Ocyclohexane, 1.4-didecyl- 76 76 32.405 1.11 C19H2802 m-Toluic acid, undec-2-enyl ester 77 77 32.525 1.25 C12H22N202 1.8-Diazeyclotetradecane-2.9-dione 90 78 32.625 0.73 C22H4602 Ethanol, 2-(eicosyloxy)- 81 81 33.194 1.84 C1BH37N0 Octadecenamide, (2)- 88 81 33.194 3.84 C1BH37N0 Octadecenamide 86 83 33.45 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.625 1.27 C2DH39N0 9-Octadecenamide, N-dimethyl- 77 85 33.790 0.56<	71	31.647	1.14	C13H27N0	N-Methyldodecanamide	81
73 31.869 2.71 C11H18N202 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(2-methylpropyl)- 62 74 32.093 2.14 C1BH3804 Triethylene glycol monododecyl ether 88 75 32.265 1.33 C26H52 Cyclohexane, 1,4-didecyl- 76 76 32.205 1.25 C12H22N202 1.8-Diazacyclotetradecane-2,9-dione 90 78 32.625 0.73 C22H4602 Ethanol, 2-(eicosyloxy)- 91 79 32.725 0.81 C18H1802 4,4-Diallyoxdiphenyl 55 80 33.050 11.86 C18H37N0 Octadecanamide 95 81 33.144 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.645 0.57 C13H27N0 N-Methyldodecanamide 77 83 33.445 0.56 C18H3802 Cyclopropaneoctanoic acid, 2[2](-2:ethylcyclopropyl)methyl]cyclopropyl]methyl], methyl ester 78 84 33.671 0.74 C2H3020 Cyclopropaneoctanoic acid, 2[2](-2:ethylcyclopropyl)methyl]cyclopropyl]methyl], methyl est	72	31.741	2.07	C15H16O2	Phenol, 4,4'-(1-methylethylidene)bis-	75
74 32.093 2.14 C18H3804 Triethylene glycol monododcyl ether 88 75 32.265 1.33 C26H52 Cyclobexae, 1.4-didecyl- 76 76 32.405 1.11 C19H2802 m-Toluic acid, undec-2-enyl ester 77 77 32.525 1.25 C12H22N202 1.8-bizacyclotetradecane-2.9-dione 90 78 32.625 0.73 C22H4602 Ethanol, 2-(eicosyloxy)- 91 79 32.725 0.81 C18H35N0 9-Octadecenamide, (Z)- 88 81 33.144 3.84 C18H37N0 Octadecenamide, (Z)- 88 82 33.370 1.17 C14H16N202 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)- 79 83 33.445 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.625 1.27 C20H39N0 9-Octadecenamide, N-dimethyl- 77 85 33.790 0.56 C2H3802 Cyclopropaneoctanic acid, 2-[[2-(2-ethylcyclopropyl]methyl], methyletert 78 86	73	31.869	2.71	C11H18N2O2	Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(2-methylpropyl)-	62
75 32.265 1.33 C26H52 Cyclohexane, 1.4-didecyl- 76 76 32.405 1.11 C19H2802 m-Toluic acid, undec-2-enyl ester 77 77 32.525 1.25 C12H22N202 1.8-biazacyclottradecane-2.9-dione 90 78 32.625 0.73 C22H4602 Ethanol, 2(eicosyloxy)- 91 79 32.725 0.81 C18H1802 A-Diallyloxydiphenyl 55 80 33.050 11.86 C18H35N0 9-Octadecenamide, (2)- 88 81 33.194 3.84 C18H37N0 Octadecenamide, NAdmen, Nethyldodecanamide 96 82 33.370 1.17 C14H16N202 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3(phenylmethyl)- 77 83 33.454 1.57 C13H27N0 N-Methyldodecanamide 88 84 33.674 0.56 C2H3802 Cyclopropaneoctanoic acid, 2-[[2-(2-ethylcyclopropyl)methyl]cyclopropyl]methyl], methyl ester 78 86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 81 <	74	32.093	2.14	C18H38O4	Triethylene glycol monododecyl ether	88
76 32.405 1.11 C19H2802 m-Toluic acid, undec-2-enyl ester 77 77 32.525 1.25 C12H22N202 1.8-Diazacyclotetradecane-2,9-dione 90 78 32.625 0.73 C22H4602 Ethanol,2-{(eiosyloxy)- 91 78 32.625 0.73 C22H4602 Ethanol,2-{(eiosyloxy)- 81 80 33.050 11.86 C18H35N0 9-Octadecenamide, (Z)- 88 81 33.194 3.84 C18H37N0 Octadecenamide, N-4(-dione, hexahydro-3-(phenylmethyl)- 79 83 33.445 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.625 1.27 C2H3802 Cyclopropaneoctanoic acid, 2-[[2-(2-ethylcyclopropyl)methyl-[cyclopropyl]methyl-, methylester 78 85 33.674 0.56 C2H3802 Cyclopropaneoctanoic acid, 2-[[2-(2-ethylcyclopropyl]methyl-[cyclopropyl]methyl-, methylester 78 86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 89 87 34.051 0.44 C16H3403 Distryl	75	32.265	1.33	C26H52	Cyclohexane, 1,4-didecyl-	76
77 32.525 1.25 C12H22N202 1,8-Diazacyclotetradecane-2,9-dione 90 78 32.625 0.73 C22H4602 Ethanol, 2-(eicosyloxy)- 91 79 32.725 0.81 C18H1802 C4.4/4-Diallyloxydiphenyl 55 80 33.050 11.86 C18H35N0 9-Octadecenamide, (Z)- 88 81 33.144 C18H37N0 Octadecanamide 88 82 33.370 1.17 C14H16N202 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)- 79 83 33.445 0.57 C13H27N0 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)- 77 84 33.625 1.27 C20H39N0 9-Octadecenamide, N-dimethyl- 77 85 33.790 0.56 C22H3802 Cyclopropaneoctanoic acid, 2-[[2-(e-thylcyclopropyl)methyl]cyclopropyl]methyl-remethyleser 78 86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 89 87 34.051 0.74 C20H4205 Tetraethylene glycol monododecyl ether 87 <td>76</td> <td>32.405</td> <td>1.11</td> <td>C19H28O2</td> <td>m-Toluic acid, undec-2-enyl ester</td> <td>77</td>	76	32.405	1.11	C19H28O2	m-Toluic acid, undec-2-enyl ester	77
78 32.625 0.73 C22H4602 Ethanol, 2-(eicosyloxy)- 91 79 32.725 0.81 C18H1802 A.4-Diallyloxydiphenyl 55 80 33.050 11.86 C18H35N0 9-Octadecenamide, (2)- 88 81 33.194 3.84 C18H37N0 Octadecenamide, (2)- 89 82 33.370 1.17 C14H16N202 Pyrrolo(1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)- 79 83 33.445 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.625 1.27 C20H39N0 9-Octadecenamide, N.N-dimethyl- 77 85 33.790 0.56 C22H3802 Cyclopropaneoctancic acid, 2-[[2-(2-ethylcyclopropyl)methyl]cyclopropyl]methyl-methylethyl esiter 78 86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 89 87 34.051 0.74 C24H500 Tetraethylene glycol monododecyl ether 91 88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 87	77	32.525	1.25	C12H22N2O2	1,8-Diazacyclotetradecane-2,9-dione	90
79 32.725 0.81 C18H1802 4.4-Diallyloxydiphenyl 55 80 33.050 11.86 C18H35N0 9-Octadecenamide, (Z)- 88 81 33.194 3.84 C18H37N0 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)- 79 82 33.370 1.17 C14H16N20 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)- 79 83 33.454 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.625 1.27 C20H39N0 9-Octadecenamide, N-M-dimethyl- 77 85 33.790 0.56 C22H3802 Cyclopropaneoctanoic acid, 2-[[2-[(2-ethylcyclopropyl)methyl]pyrolpyl]methyl]-methylethyl est 78 86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 89 87 34.051 0.74 C24H500 Tetracthylene glycol monododecyl ether 91 88 34.574 0.87 C20H4205 Tetracthylene glycol monododecyl ether 91 89 34.721 1.01 C10H1803 1,3-Dioxolan-4-one	78	32.625	0.73	C22H46O2	Ethanol, 2-(eicosyloxy)-	91
80 33.050 11.86 C18H35N0 9-Octadecenamide, (Z)- 88 81 33.194 3.84 C18H37N0 Octadecanamide 95 82 33.370 1.17 C14H16N202 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl]- 79 83 33.445 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.625 1.27 C20H39N0 9-Octadecenamide, N,N-dimethyl- 77 85 33.700 0.56 C22H3802 Cyclopropaneoctancia acid, 2-[[2-{(2-ethylcyclopropyl)methyl]cyclopropyl]methyl- methyl ester 78 866 33.874 0.56 C18H3604 Triethylene glycol monododecyl ether 89 87 34.051 0.74 C20H4205 Tetraethylene glycol monododecyl ether 91 88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 87 90 35.292 0.44 C16H3403 Diethylane glycol monododecyl ether 87 91 36.075 0.21 C36H58 Tisthylane glycol monododecyl ether <t< td=""><td>79</td><td>32.725</td><td>0.81</td><td>C18H18O2</td><td>4,4'-Diallyloxydiphenyl</td><td>55</td></t<>	79	32.725	0.81	C18H18O2	4,4'-Diallyloxydiphenyl	55
81 33.194 3.84 C18H37N0 Octadecanamide 95 82 33.370 1.17 C14H16N202 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)- 79 83 33.445 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.625 1.27 C20H39N0 O-9-Octadecenamide, NN-dimethyl- 77 85 33.874 0.56 C22H3802 Cyclopropaneoctanoic acid, 2-[[2-(2-ethylcyclopropyl)methyl]cyclopropyl]methyl]-, methyl ester 78 86 33.874 0.56 C18H3804 Trietnylene glycol monododecyl ether 89 87 34.051 0.74 C20H4205 Tetraethylene glycol monododecyl ether 91 88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 87 90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontatetrayne 57 92 36.154 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl	80	33.050	11.86	C18H35NO	9-Octadecenamide, (Z)-	88
82 33.370 1.17 C14H16N202 Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)- 79 83 33.445 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.625 1.27 C20H39N0 9-Octadecenamide, N,N-dimethyl- 77 85 33.790 0.56 C22H3802 Cyclopropaneoctanoic acid, 2-[[2-(2-ethylcyclopropyl)methyl]cyclopropyl]methyl]-, methyl ester 78 86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 89 87 34.051 0.74 C24H500 N-Methylene glycol monododecyl ether 91 88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 91 89 34.571 1.01 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl), (2S-cis)- 75 90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl), (2S-cis)- 75 92 36.154 0	81	33.194	3.84	C18H37NO	Octadecanamide	95
83 33.445 0.57 C13H27N0 N-Methyldodecanamide 88 84 33.625 1.27 C20H39N0 9-Octadecenamide, N,N-dimethyl- 77 85 33.790 0.56 C22H3802 Cyclopropaneoctanoic acid, 2-[[2-[(2-ethylcyclopropyl)methyl]cyclopropyl]methyl]-, methyl ester 78 86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 89 87 34.051 0.74 C24H500 n-Tetracosanol-1 83 88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 91 89 34.721 1.01 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl)-, (2S-cis)- 75 90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontaterayne 57 92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 88 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethyl	82	33.370	1.17	C14H16N2O2	Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(phenylmethyl)-	79
84 33.625 1.27 C20H39N0 9-Octadecenamide, N,N-dimethyl- 77 85 33.790 0.56 C22H3802 Cyclopropaneoctanoic acid, 2-[[2-((2-ethylcyclopropyl)methyl]cyclopropyl]methyl]-, methyl ester 78 86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 89 87 34.051 0.74 C24H500 n-Tetracosanol-1 83 88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 91 89 34.721 1.01 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5-(1-methylethyl)-, (2S-cis)- 75 90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontatetrayne 57 92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 90 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5 (1-methylethyl)-, (2S-cis)- 75 94 36.933 0.64 C18H3407	83	33.445	0.57	C13H27N0	N-Methyldodecanamide	88
85 33.790 0.56 C22H3802 Cyclopropaneoctanoic acid, 2-[[2-[(2-ethylcyclopropyl)methyl]cyclopropyl]methyl]-, methyl ester 78 86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 89 87 34.051 0.74 C24H500 n-Tetracosanol-1 83 88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 91 89 34.721 1.01 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5-(1-methylethyl)-, (2S-cis)- 75 90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontatetrayne 57 92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 90 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5-(1-methylethyl)-, (2S-cis)- 75 94 36.933 0.64 C18H3407 2,2'-(2,2'Oxybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 <td>84</td> <td>33.625</td> <td>1.27</td> <td>C20H39NO</td> <td>9-Octadecenamide, N,N-dimethyl-</td> <td>77</td>	84	33.625	1.27	C20H39NO	9-Octadecenamide, N,N-dimethyl-	77
86 33.874 0.56 C18H3804 Triethylene glycol monododecyl ether 89 87 34.051 0.74 C24H500 n-Tetracosanol-1 83 88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 91 89 34.721 1.01 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5-(1-methylethyl), (2S-cis)- 75 90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontatetrayne 57 92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 90 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5-(1-methylethyl), (2S-cis)- 75 94 36.933 0.64 C18H3407 2,2'-(2,2'-Oxybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39N0	85	33.790	0.56	C22H38O2	Cyclopropaneoctanoic acid, 2-[[2-[(2-ethylcyclopropyl)methyl]cyclopropyl]methyl]-, methyl ester	78
87 34.051 0.74 C24H500 n-Tetracosanol-1 83 88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 91 89 34.721 1.01 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5-(1-methylethyl), (2S-cis)- 75 90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontatetrayne 57 92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 90 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5-(1-methylethyl)-, (2S-cis)- 75 94 36.933 0.64 C18H3804 Z/2'(2,2'-Oxybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39N0 Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408	86	33.874	0.56	C18H38O4	Triethylene glycol monododecyl ether	89
88 34.574 0.87 C20H4205 Tetraethylene glycol monododecyl ether 91 89 34.721 1.01 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl)-, (2S-cis)- 75 90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontatetrayne 57 92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 90 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl)-, (2S-cis)- 75 94 36.933 0.64 C18H3407 2,2'-(2,2'-Oxybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39NO Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 <	87	34.051	0.74	C24H500	n-Tetracosanol-1	83
89 34.721 1.01 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl)-, (2S-cis)- 75 90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontatetrayne 57 92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 90 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl)-, (2S-cis)- 75 94 36.933 0.64 C18H3407 2,2'-(2,2'Oxybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39NO Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41	88	34.574	0.87	C20H42O5	Tetraethylene glycol monododecyl ether	91
90 35.292 0.44 C16H3403 Diethylene glycol monododecyl ether 87 91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontatetrayne 57 92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 90 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl)-, (2S-cis)- 75 94 36.933 0.64 C18H3407 2,2-(2,2-Oxybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39N0 Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41 C30H61Br Triethylene glycol monododecyl ether 72 100 39.985 1.02 C27H44 Cholesta-3	89	34.721	1.01	C10H18O3	1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5-(1-methylethyl)-, (2S-cis)-	75
91 36.075 0.21 C36H58 15,17,19,21-Hexatriacontatetrayne 57 92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 90 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl)-, (2S-cis)- 75 94 36.933 0.64 C18H3407 2,2'-(2,2'-0xybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39NO Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41 C30H61Br Triacontane, 1-bromo- 72 100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	90	35.292	0.44	C16H34O3	Diethylene glycol monododecyl ether	87
92 36.154 0.43 C18H3804 Triethylene glycol monododecyl ether 90 93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl)-, (2S-cis)- 75 94 36.933 0.64 C18H3407 2,2'-(2,2'-0xybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39NO Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41 C30H61Br Triacontane, 1-bromo- 72 100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	91	36.075	0.21	C36H58	15,17,19,21-Hexatriacontatetrayne	57
93 36.838 0.43 C10H1803 1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5(1-methylethyl)-, (2S-cis)- 75 94 36.933 0.64 C18H3407 2,2'-(2,2'-Oxybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39NO Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41 C30H61Br Triacontane, 1-bromo- 72 100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	92	36.154	0.43	C18H38O4	Triethylene glycol monododecyl ether	90
94 36.933 0.64 C18H3407 2,2'-(2,2'-Oxybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate 75 95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39NO Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41 C30H61Br Triacontane, 1-bromo- 72 100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	93	36.838	0.43	C10H18O3	1,3-Dioxolan-4-one, 2-(1,1-dimethylethyl)-5-(1-methylethyl)-, (2S-cis)-	75
95 37.136 0.30 C20H4205 Tetraethylene glycol monododecyl ether 88 96 37.636 0.31 C22H39N0 Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41 C30H61Br Triacontane, 1-bromo- 72 100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	94	36.933	0.64	C18H3407	2,2'-(2,2'-Oxybis(ethane-2,1-diyl)bis(oxy))bis(ethane-2,1-diyl) dipentanoate	75
96 37.636 0.31 C22H39NO Pyrrolidine, 1-(1-oxo-10-octadecynyl)- 66 97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41 C30H61Br Triacontane, 1-bromo- 72 100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	95	37.136	0.30	C20H42O5	Tetraethylene glycol monododecyl ether	88
97 38.272 0.57 C26H5408 Heptaethylene glycol monododecyl ether 92 98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41 C30H61Br Triacontane, 1-bromo- 72 100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	96	37.636	0.31	C22H39NO	Pyrrolidine, 1-(1-oxo-10-octadecynyl)-	66
98 39.542 0.54 C18H3804 Triethylene glycol monododecyl ether 88 99 39.835 0.41 C30H61Br Triacontane, 1-bromo- 72 100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	97	38.272	0.57	C26H54O8	Heptaethylene glycol monododecyl ether	92
99 39.835 0.41 C30H61Br Triacontane, 1-bromo- 72 100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	98	39.542	0.54	C18H38O4	Triethylene glycol monododecyl ether	88
100 39.985 1.02 C27H44 Cholesta-3,5-diene 94	99	39.835	0.41	C30H61Br	Triacontane, 1-bromo-	72
	100	39.985	1.02	C27H44	Cholesta-3,5-diene	94

the liquid fraction are compounds containing nitrogen or nitrogen and oxygen atoms in their structure. This finding is consistent with studies by Marcilla et al. (2012) and Kluska et al. (2019), which also identified nitrogenated compounds as the main group of compounds from the analysis of liquid fraction from leather pyrolysis. The percentage of nitrogenated compounds in this study were, however, significantly lower in comparison to the study by Marcilla. The justification for this difference is because this study utilized slow pyrolysis in comparison to fast pyrolysis in the study by Marcilla et al. (2012). This is an important factor to consider as it influences the liquid product composition. The second major group found in the liquid fraction was oxygenated compounds (containing ketones, aldehydes, alcohols, and acids), followed by aromatics, phenols, alkanes, and alkenes, and lastly, sulfur-containing compounds. The

compound distribution found in this study exhibit similarities to those reported by Fang et al. (2018).

Based on the composition of the liquid fraction, it would be useful for the following applications. The abundance of NH₂ groups present in the liquid fraction makes it potentially valuable for the production of fertilizers (Marcilla et al., 2012). According to Yilmaz et al. (2007), the liquid yield can additionally be useful as a chemical feedstock or a fuel after retreatments such as hydrogenation, Fischer–Tropsch synthesis, or stream cracking. Some of the main compounds from the liquid chemistries including Octadecanamide and 13-Docosenamide have useful applications as adhesives and sealant chemicals, anti-adhesive agents, as well as lubricants and or surface agents (NCBI, 2020a., and NCBI, 2020b). According to Getachew et al. (2016), oleamides may present industrial applications such as lubricants,

TABLE 11: Liquid yield compound classification.

Classification type	Percentages of compounds (%)
N or N/O compounds	40
O compounds	35
Alkanes and alkenes	9
Aromatics	6
Phenols	6
S compounds	2
	100

(N or N/O = compounds containing N or N and O; O compounds = compounds containing oxygen; alkanes and alkenes=hydrogen and carbon compounds; aromatics = aromatics; phenols = phenol and its derivates; S compounds =compounds containing sulfur)

corrosion inhibitors and slip agents. Specific compounds, such as pyrrole and its derivatives, can be useful as intermediates in the synthesis of pharmaceuticals, perfumes, agrochemicals, and dyes (Marcilla et al., 2012, and Kaur et al., 2017). To achieve the applications for leather footwear pyrolysis products it is necessary to consider that processing would be required in order to improve the product quality through purifying, refining and upgrading the products.

A limitation of pyrolysis in this study, and in general, is that it relies on an external energy supply to reach its processing temperatures, which is often derived from non-renewable sources. Future leather pyrolysis studies should therefore consider solar assisted pyrolysis to overcome this obstacle by prioritizing the utilization of solar energy for thermochemical processing (Morales et al., 2014, and Weldekidan et al., 2018).

4. CONCLUSIONS

This study investigated whether leather from post-consumer footwear could be pyrolyzed using gram-scale (fixed-bed) and microgram-scale (TGA) pyrolysis reactors. The distribution of the product fractions proved that leather footwear pyrolysis was viable under the given conditions. It was shown through an examination of pyrolysis product distribution and characterization that leather footwear has waste recovery potential, hence contributing to the overall aim of the study.

The results of this study showed that pyrolysis temperature has a significant effect on the product yields. According to the results obtained, the liquid yield was maximized within the temperature range of 550-650°C (Max= 54 wt.%). The solid yield decreased substantially from 43 wt.% to 25 wt.% with an increase in the reactor temperature. The solid yield indicated the completion of leather footwear pyrolysis at 650°C since a constant value of approximately 25 wt.% was reached. This was verified since the solid yield remaining corresponded to the fixed carbon and ash proportions characterized in the leather sample. There was a general increase of gas yields as a function of higher temperatures and longer residence times, explained by decreasing char yields in the leather sample.

The pyrolysis product yields were characterized, and the char yield revealed that it may be suitable as a solid fuel due to its high calorific value (25.55 MJ/kg). A wide distribution of pyrolytic compounds were presented in this study, with nitrogenated derivatives forming the major group from the liquid yield. Based on the composition of the liquid yields, potential applications include use in the production of fertilizers, use as a chemical feedstock and use in the pharmaceutical industry. In addition, some of the main compounds found present potential industrial applications including use as adhesives, sealant chemicals, anti-adhesive agents, lubricants, and surface agents. Due to the composition of the pyrolysis product yields, however, it is necessary to consider retreatment before application to remove impurities. The pyrolysis oil presented a gross calorific value higher than biomass derived pyrolysis oils and low-grade coal. This indicated its potential as an energy source or fuel. It can therefore be concluded that the leather pyrolysis oils present a more valuable source of energy than some biomass pyrolysis oils.

Future work should investigate the quantitative aspect of the compounds present in the pyrolysis yields. This would further substantiate the potential for reuse applications and the viability of recovering these materials from leather footwear in practice.

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